ELECTROPHORETIC PROTEIN PATTERNS OF FARM GROWN AND WILD EDIBLE OYSTER CRASSOSTREA MADRASENSIS FROM DIFFERENT LOCATIONS

P. PHILIP SAMUEL AND R. THANGAVELU*

CRME Field Station (ICMR), Periyar Nagar, Virudhachalam 606 001

ABSTRACT

Electrophorograms of adductor muscle, mantle and gill of farm grown and wild edible oyster C. madrasensis exhibited distinct differences. Marked protein pattern variations were also evident between the oysters of Ennore Estuary and Muttukadu Backwater near Madras in Tamil Nadu, India. Tissue, environment and locality specific differences have been discussed in the light of number, thickness and staining densities of protein fractions.

INTRODUCTION

AQUATIC organisms present a striking challenge for applied genetics. Most cultivable species still lack information of their genotype and phenotype. Electrophoretic studies of serum protein, muscle protein, plasma protein and haemoglobin, especially of higher vertebrates have revealed species specific pattern (Connell. 1953; Tsuyuki et al., 1962). Gene variant patterns with frequencies characteristic of particular geographic areas or races have been observed in Batevgobius and Pomacentridae (Gorman et al., 1976; Gorman and Kim, 1977). Here an attempt has been made to find out the regional variation of protein pattern of commercially important edible oyster Crassostrea madrasensis.

The authors are grateful to Dr. M. J. George for his help and suggestions.

MATERIAL AND METHODS

Oysters for the present study were collected from three locations, namely the natural beds of Ennore. Muttukadu and Muttukadu oyster farm. The oysters were shucked and the meat was thoroughly washed with cold double distilled water and then blotted dry. The mantle, gill and adductor muscle were dissected out. One gram of each tissue was homogenised with ice cold double distilled water. The homogenate was centrifuged for 15 minutes at 3000 rpm. The supernatant containing dissolved water soluble proteins was used as the sample for electrophoresis. Polyacrylamide gel (7%) electrophoresis was performed. making use of the procedure of Laemmli (1970) at 4°C, 240 volts, 48 mA for 31 hours. After completion of electrophoresis. gels were stained with coomassie Brilliant Blue. Gels were destained and stored in 7% acetic acid. They were photographed and scanned in ultrascanner.

RESULTS

The results of 17 experiments carried out on tissues from the mantle, gill and adductor

[•] Present address : Madras Rescarch Centre of CMFRI, 141, Marshalls Road, Madras-600 008.

muscle of the edible oyster on the electrophoretic protein fractions have been assigned numbers by keeping in mind the number of crests found to correspond to the number of distinct proteins and the areas under the crests proportional to their concentrations.

M antle

The mantle tissue of O. madrasensis of Ennore Estuary exhibited ten protein fractions of which 3 bands were thick 3 thinner and others were minor bands whereas in the farm bred oyster 5. 6, 7 & 8 th fractions were absent there being only six bands and in Muttukadu specimens 7 th and 8 th fractions were absent there being only eight bands (Fig. 1). Thus six protein bands were common for all the

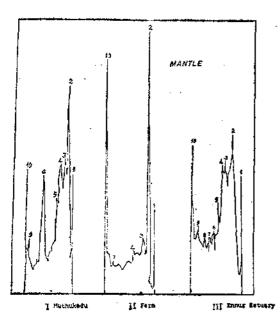


Fig. 1. Electrophorogram of mantle tissue of edible oyster from Muttukadu, Muttukadu Farm and Ennore Estuary.

oysters obtained from the three different localities. The remining 4 smaller or thinner bands showed variations or altogether absent. Gill

. C. madrasensis of Ennore and farm area expressed eleven protein fractions in the gills tested. In Muttukadu C. madrasensis specimens, fractions 6, 8 and 9 were absent. Totally eight common bands were seen in the gill of C. madrasensis of different areas tested (Fig. 2).

Adductor muscle

Totally ten protein fractions were observed in the adductor muscle of C. madrasensis (Fig. 3). All the ten protein fractions were observed in the muscle of C. madrasensis of farm area. In the muscle of Ennore Estuary area specimens 6th, 8th and 9th fractions were absent and in C. madrasensis of Muttukadu 5th and 7th bands were absent. There were common six bands of muscle,

Environmental parameters

The environmental parameters such as salinity, temperature and dissolved oxygen and their annual range in the three different places are shown in Table 1.

 TABLE 1. Environmental parameters of three different localities

Place	Salinity (‰)	Temperature (°C)	Dis. oxygen (ml/l)
Ennore	14.52-33.4	28.0-34.0	3.52-5.24
Muttukadu	13,86-39,7	28.1-34.0	3.14-5.82
Muttukadu Farm	6.32-36,0	28.5-34.5	3.00-5,30

DISCUSSION

The edible oyster *C. madrasensis* is widely distributed on the east and west coasts of India. Several view points have been expressed regarding the similarities and dissimilarities of protein patterns of the same species occurring in different localities. Geographical variations in biochemical and serological tests have been reported due to a variety of physiological processes, activities and tolerances by Menzel (1956), Numachi (1962) and Hilman (1964). Galtsoff (1964) however, observed that the shell ing the bio-chemical genetics of the blue gill Lepomis macrochirus, has suggested that the reason for this type of variation may be the discontinuous nature of aquatic habitat. Janson

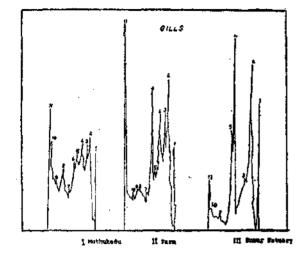


Fig. 2. Electrophorogram of gill tissue of edible oyster from Muttukadu, Muttukadu Farm and Ennore Estuary.

morphology was being influenced by local environmental conditions, but did not report any consistent variation among geographically distinct populations.

The present electrophoretic study of the different tissues of the same species from areas suggests that the samples from Ennore, Muttukadu and oyster farm possess distinct protein patterns in terms of the number of bands and also in presence or absence of certain protein fractions. These variations may be brought about by the differences in the environment especially salinity and exposure of the natural beds during low tide, and the resultant physiological stresses. Avise and Smith (1984) study-

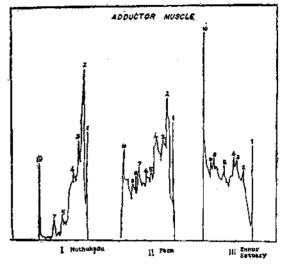


Fig. 3. Electrophorogram of adductor muscle of edible oyster from Muttukadu, Muttukadu Farm and Ennore Estuary.

and Ward (1984) has mentioned that the differences in populations occupying different areas separated by physical barriers could be related directly to environmental pressure like exposure during low tide and to substrata occupied by them. A heterogenous environment would enhance genetic variation and a homogenous environment erode variation according to Fujiyo *et al.* (1983).

The results of the present study suggest that the natural oyster beds of *C. madrasensis* which lie in the intertidal area being subjected to tidal amplitude and also facing a complex of environmental changes undergo considerable physiological stresses and this has resulted in the protein band variations as reported here.

References

AVISE, J. C. AND M. H. SMITH 1984. Biochemical genetics of Sunfish: Geographic variation and subspecific intergraduation in the blue gill Lepomis macrochirus. Evolution, 28: 42-56.

CONNELL, J. J. 1953. Studies on the proteins of fish skeletal muscle. 2. Electrophoretic analysis of low ionic strength extracts of several species of fish. *Biochem. J.*, 55: 328-388.

FURYO YOS HIHLS, RYOICHI YAMANAKA AND P. J. SMITH 1983. Genetic variation in marine molluscs. Bull, Jap. Soc. Scientific fishing, 49 (12): 1809-1817.

GALTSOFF, P. S. 1954. The American oyster Crassostrea virginica (Gmelin). Fishery Bulletin of the Fish and Wildlife service, 64. U. S. Government Printing Office, Washington, 25: 480.

GORMAN, C. G. AND Y. K. KIM 1977. Genotypic evolution in the face of phenotypic conservativeness: Abudefdof (Pomacentriade) from the Atlantic and Pacific sides of Panama. *Copeia*, 694-697.

GORMAN C. C., Y. J. KIM AND R. RUBINOFF 1976. Genetic relationships of three species of *Bathygobius* from the Atlantic and Pacific sides of Panama. *Ibid.*, 361-364. HILLMAN, R. E. 1964. Chromatographic evidence of intraspecific genetic differences in the eastern oyster Crassostrea viriginica. Systematic Zoology, 13: 12-18.

JANSON K. AND R. D. WARD 1984. Microgeographic variation in allozymes and shell characters in *Littorine* saxatilis Olivi (Prosobranchia : Littorinidae). Biological and Linn. Soc., 22: 289-307.

LAEMMLI, U. K. 1970. Cleavage of structural proteins during the assembly of the head of bactriophage T-4. Nature, 227: 680-688.

MENZEL, R. W. 1956. The effect of temperature on the ciliary action and other activities of oysters. *Florida State University studies*, 22: 25-36.

NUMACHI, K. 1962. Serological studies of species and races of oysters. American Naturalist, 96: 211-217.

TSUYUKI, H. E. ROBERTS AND R. E. A. CADD 1962. Muscle proteins of Pacific Salmon (Oncorhynchus) III. The separation of muscle protein soluble in low ionic strength salt solutions by starch gel electrophoresis. Canadian J. Biochem. and Physiol., 40: 929-936.